









Table I. Microwave Data for Propylene Oxide Isotopomeric Mixtures

run	transition, peak intensity (trans- d_1/d_0)					$n_{t-d_1}/n_{d_0}^a$
	9 _{2,7} -9 _{1,8}	10 _{2,8} -10 _{1,9}	11 _{2,9} -11 _{1,10}	12 _{2,10} -12 _{1,11}	13 _{2,11} -13 _{1,12}	
2	38/9	41/11	54/12	66/16	81/18	4.8 (0.3)
3	17/3	21/4.5	26.5/5	32/7	40/8	5.7 (0.5)
4	0.9/15	0.9/17.5	1.1/20	1.6/28	1.8/30	0.06 (0.01)
5	0.95/21	1.11/25	1.3/31	1.6/41	2.2/48	0.05 (0.01)
6	15.5/7	19/8	21.5/10	30/12	35/14	2.5 (0.2)
7	3.0/24	3.0/29	3.5/31	4/42	5/46	0.12 (0.01)
8	24/6.8	30/7.9	37/9.6	47/11.9	61/14	4.4 (0.2)
9 ^b	17/13	24/14	30/18	39/23	48/29	1.6 (0.2)

^a Average isotopomer ratio and standard deviation calculated following ref 5a; for each transition $n_a/n_b = R^{-1}(I_a/I_b)$ where I_a is the peak intensity for species a etc. and R converts intensity ratios into the mole ratio n_a/n_b . The values of R for the five transitions are respectively 0.86, 0.87, 0.88, 0.88, and 0.89 for n_{t-d_1}/n_{d_0} and 1.0, 0.99, 0.97, 0.96, and 0.95 for n_{t-d_1}/n_{c-d_1} . ^b Peak intensities for trans- $d_1/cis-d_1$; d_0 not determined.

Table II. Epoxidation of Propylene by Cytochrome P-450_{LM2}

run	substrate	conditions	epoxide yield, μmol	% d_0	% trans- d_1
1		P-450/reductase/H ₂ O, NADPH/O ₂	1.6	100	
2		P-450/Reductase/D ₂ O, NADPH/O ₂	2.2	17	83 ^b
3		P-450/Reductase/H ₂ O, NADPH/O ₂	2.4 ^a	15	85
4		P-450/Reductase/H ₂ O, NADPH/O ₂	2.3	94	6 ^b
5		P-450/ ϕ IO/H ₂ O	1.8	95	5 ^b
6		P-450/Reductase/D ₂ O, NADPH 3 days	2.0	27	73 ^b
7		Fe(TPP)Cl/Ph-IO	60	18	82 ^b
8		Mn(TPP)Cl/Ph-IO	50	38 ^f	62

^a Recovered propylene showed 4.7% trans-1-deuteriopropylene. ^b Less than 1% cis-1-deuteriopropylene oxide was detected. Loss of deuterium was confirmed by GC-mass spectrometry (m/e 58, 43). ^c Starting material was 72% trans-1-deuteriopropylene, 28% propylene. ^d Deuterium content of recovered propylene was unchanged. ^e Starting material was 80% trans-1-deuteriopropylene, 20% propylene. ^f cis-1-Deuteriopropylene oxide; propylene oxide not determined.

species is an attractive candidate for the "active oxygen species" in the cytochrome P-450 cycle. Although the detailed mechanism of olefin epoxidation by simple oxometal species is not yet clear, the interesting suggestion that chromyl reagents⁸ and oxomanganese porphyrins⁹ react with carbon-carbon double bonds to form an oxametallacycle may offer an opportunity for proton exchange. Thus, the cycloaddition of propylene with **1** could reasonably form **2**. There is ample precedent that metal alkyls can be deprotonated to form metal carbenes.¹⁰ Loss of a proton from **2** would provide access to the iron carbene **3**. Metallo-carbene species have been suggested to result from the reductive metabolism of halocarbons by cytochrome P-450.¹¹ Further, an iron carbene such as **3** would provide a path to the *N*-alkylporphyrins **4** which are known to derive from terminal alkenes.¹²

(8) Sharpless, K. B.; Teranishi, A. Y.; Bäckvall, J. E. *J. Am. Chem. Soc.* 1977, 99, 3120.

(9) Collman, J. P.; Brauman, J. I.; Meunier, B.; Raybuck, S. R.; Koddeck, T. *Proc. Natl. Acad. Sci. U.S.A.* 3245-3248.

(10) (a) Schrock, R. *Acc. Chem. Res.* 1979, 12, 98. (b) We thank R. Grubbs for an insightful discussion of this process.

(11) (a) Mansuy, D. *Pure Appl. Chem.* 1980, 82, and references therein. (b) Mansuy, D.; Lange, M.; Chottard, J.-C. *J. Am. Chem. Soc.* 1978, 100, 3213. (c) Mansuy, D.; Battioni, J.-P.; Chottard, J.-C.; Ullrich, V. *J. Am. Chem. Soc.* 1971, 101, 3971. (d) Mansuy, D.; Lange, M.; Chottard, J.-C. *J. Am. Chem. Soc.* 1979, 101, 6437.

(12) (a) Ortiz de Montellano, P. R.; Correia, M. A. *Annu. Rev. Pharmacol. Toxicol.* 1983, 23, 481 and references cited therein. (b) Ortiz de Montellano, P. R.; Mangold, B. L. K.; Wheller, C.; Kunze, K. L.; Reich, N. D. *J. Biol. Chem.* 1983, 258, 4208. (c) Kunze, K. L.; Mangold, B. L. K.; Wheller, C.; Beilan, H. S.; Ortiz de Montellano, P. R. *J. Biol. Chem.* 1983, 258, 4202. (d) Consistent with these reports the epoxidation of 1-hexene in D₂O by reconstituted cytochrome P-450 resulted in negligible deuterium incorporation. The oxidation of 1-butene with the reconstituted P-450 system in D₂O has been shown by microwave spectroscopy and mass spectrometry to give 1-butene oxide with partial deuterium incorporation.

The observed preference for trans protonation can be accommodated in Scheme I either by a stereoselective protonation of **3** to regenerate **2** or by ring closure to form the metallooxirane **5**. Epoxide production in the former process would result from reductive elimination from **2** while protolysis of the carbon metal bond would produce the epoxide from **5**. No definitive choice between these possibilities can be made with the data at hand, but mechanisms for propylene epoxidation which do not allow for proton exchange must be eliminated on the basis of these results. Efforts to detect intermediates such as **2**, **3**, and **5** in enzymic and model porphyrin-mediated epoxidations are currently under way.¹³

Acknowledgment. Support of this research by the National Institutes of Health (GM 25923) is gratefully acknowledged. The National Science Foundation provided funds to purchase and maintain the microwave spectrometer used in this work. Purified cytochrome P-450_{LM2}, cytochrome P-450 reductase, and technical advice were generously provided by Professor M. J. Coon, Department of Biological Chemistry, The University of Michigan.

Registry No. P-450, 9035-51-2; Fe(TPP)Cl, 16456-81-8; Mn(TPP)Cl, 32195-55-4; CH₂=CHCH₃, 115-07-1; (*F*)-CHD=CHCH₃, 1560-60-7; iodosulbenzene, 536-80-1; monooxygenase, 9038-14-6; propylene oxide, 75-56-9; trans-1-deuteriopropylene oxide, 34074-743.

(13) Groves, J. T.; Watanabe, Y. *J. Am. Chem. Soc.* 1986, 108, 507-508.

Scanning Electrochemical and Tunneling Ultramicroelectrode Microscope for High-Resolution Examination of Electrode Surfaces in Solution

Hsue-Yang Liu, Fu-Ren F. Fan, Charles W. Lin, and Allen J. Bard*

Department of Chemistry, University of Texas
Austin, Texas 78712

Received March 13, 1986

We describe here a new apparatus for examination of conductor (or semiconductor) surfaces immersed in solution by measurement of either the electrochemical or tunneling current that flows when an ultramicroelectrode tip is scanned above the surface. Results with a Pt-coated test structure yield a spacial resolution of about 30 nm. Most methods for examination of solid surfaces, e.g., scanning electron microscopy (SEM) or the various electron spectroscopies, require the sample to be in an ultrahigh vacuum (UHV) environment and are not suitable for in situ electrochemical studies. Optical techniques for in situ electrode surface studies exist (e.g., infrared and surface-enhanced Raman spectroscopy) but they are capable of only limited spacial resolution, as governed by diffraction limitations and source beam size. An alternative approach was suggested by recent investigations with scanning tunneling microscopy (STM) that demonstrate the possibility of direct determination of surface structure with atomic resolution.¹⁻⁵ In this method a very sharp metal tip is brought

